



# Utilizing Biosensors to Monitor Gaping Behavior of *Crassostrea virginica* (Eastern Oyster)

Jessie Burgoyne, Danielle Slater, Adam Hookway, Selina Cheng, Cam Ragland, Easton White  
Department of Biological Sciences, University of New Hampshire, Durham, NH 03824



## Introduction

- Eastern oysters demonstrate gaping behaviors for feeding, respiration, and other physiological functions.<sup>2</sup>
- Environmental stressors can alter gaping behavior, making oysters an indicator of water quality.<sup>1</sup>
- As salinity levels in Great Bay shift throughout the year, understanding how oysters respond is important for farms and restoration projects.
- By utilizing magnetic sensors, the correlation between gaping behavior and salinity could be recorded.

## Research Goals and Questions

- How does the gaping behavior of Eastern oysters correlate with salinity, and can gaping behavior be used as an indicator of water quality?
- Modify the existing sensors to facilitate long-term deployments by establishing power-independence.

## Methods (Biology)

- 8 Oysters were attached to a sensor unit and kept in a tank within Spaulding Hall's Aquatic Core Lab.
- The biosensors recorded the duration and frequency of each oyster's gaping behavior.
- The tank was kept at optimal water quality parameters for 2 weeks before salinity was dropped from 20-25ppt to 10-15ppt. After dropping the salinity, data on gaping behavior was collected for another week.
- A separate biosensor unit with 6 oysters was deployed at NH Sea Grant's Aquafort for 2 weeks to collect continuous data on gaping behavior.



Figure 1: Tank with oysters (top), saltwater test kit (bottom left), water quality sensor (bottom middle), and labeled oysters (bottom right).

## Methods (Engineering)

- Define baseline power requirement parameters - 1.2 W (watts) with 6 sensors.
- Modify hardware and firmware to limit power needs - 0.6W.
- Double solar potential from 10W to 20W.
- Upgrade solar harvesting module from 2W to 20W.
- Increase power storage from 10 Ah (amp hours) to 50Ah.
- **Max Deployment length:**
  - Initially: ~3 days
  - Upgraded: ~2 months



Figure 2: First Power Independent Deployment on AquaFort

## Results

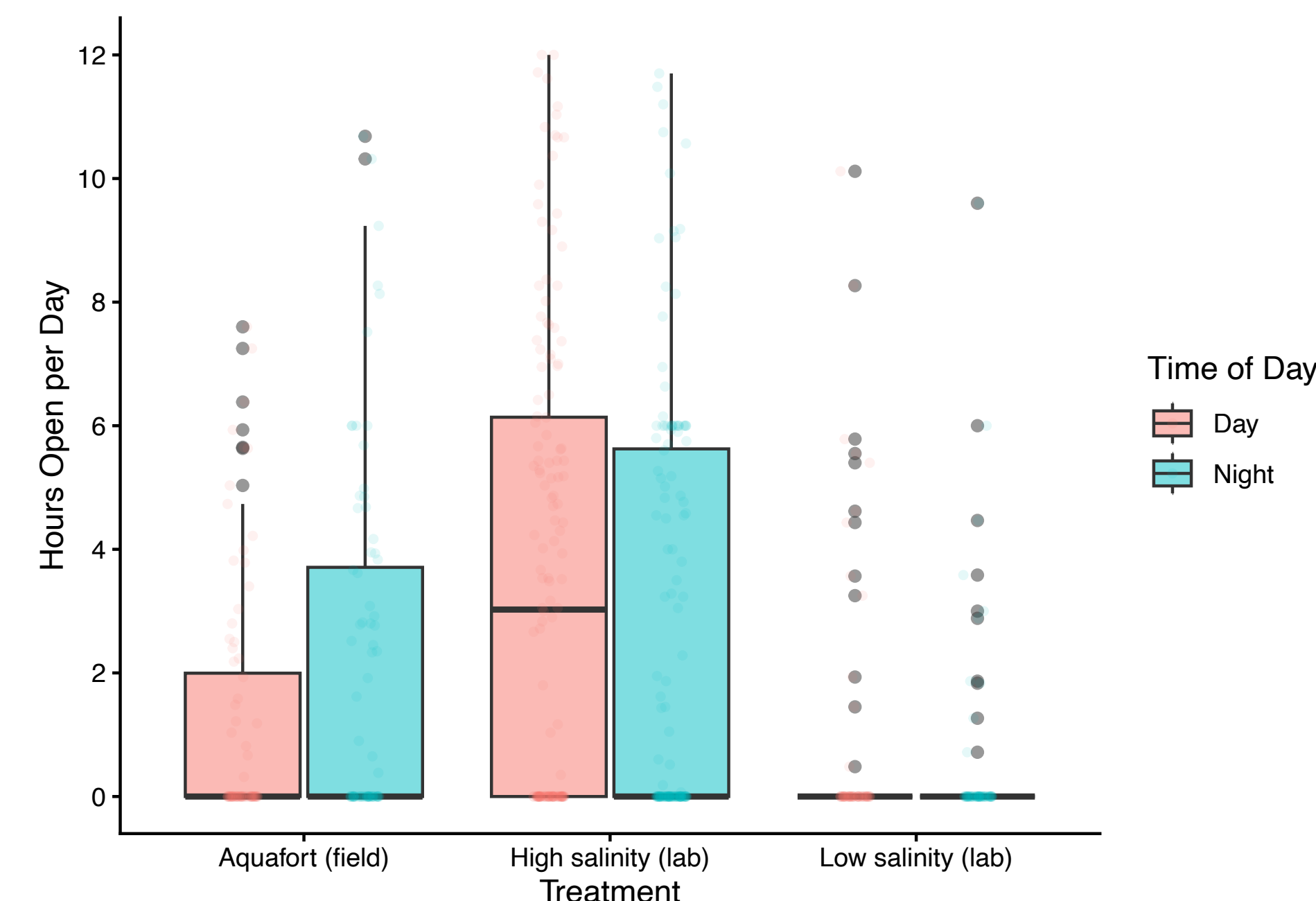


Figure 3: Hours per day oysters are open at the Aquafort and across salinity treatments in the lab experiment.

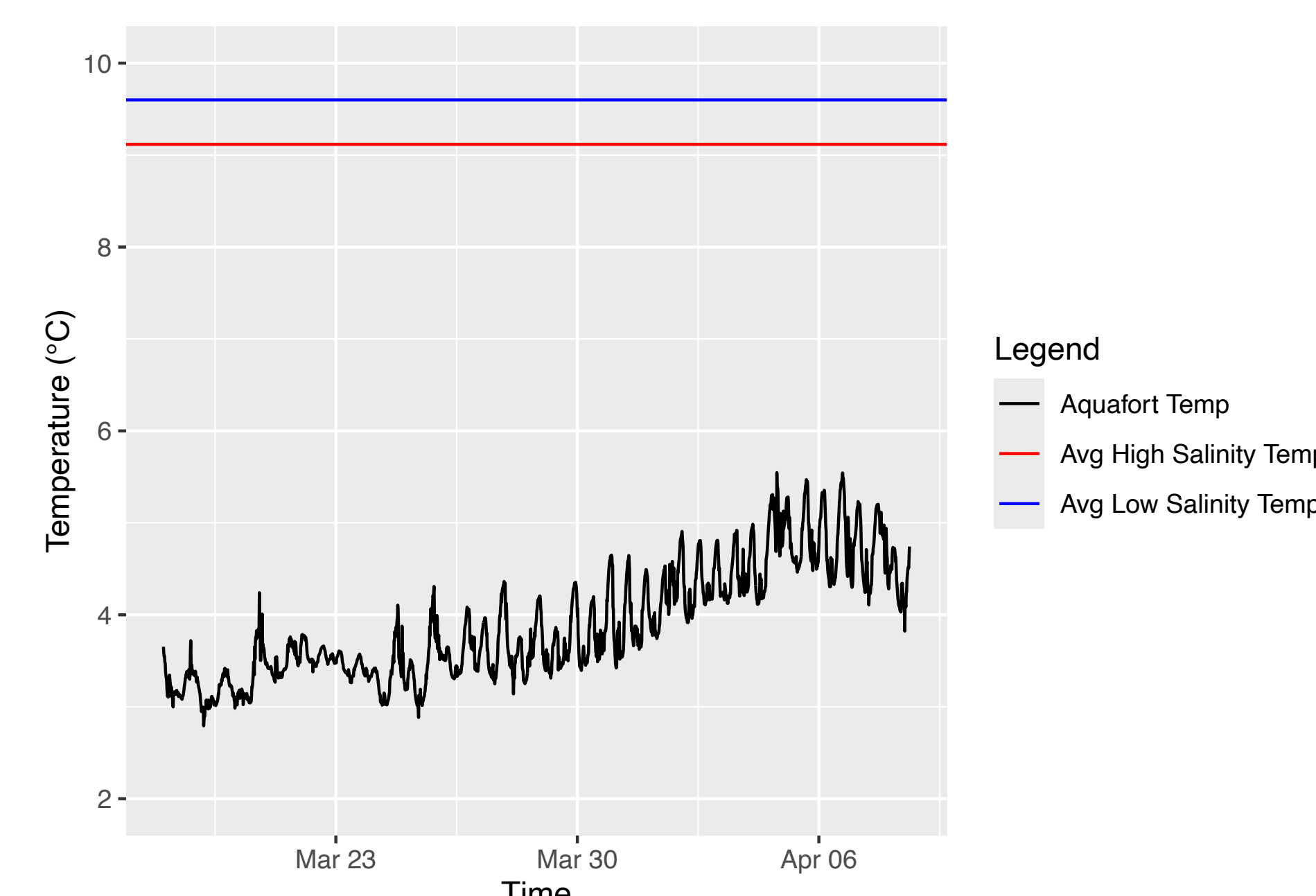


Figure 5: Water temperature at Aquafort during biosensor deployment with average temperatures of high and low salinity lab treatments

- The average temperature was 9.1°C in the high salinity treatment and 9.6°C in the low salinity treatment.
- The average salinity was 22.8 ppt in the high salinity treatment and was 13 ppt in the low salinity lab treatment.

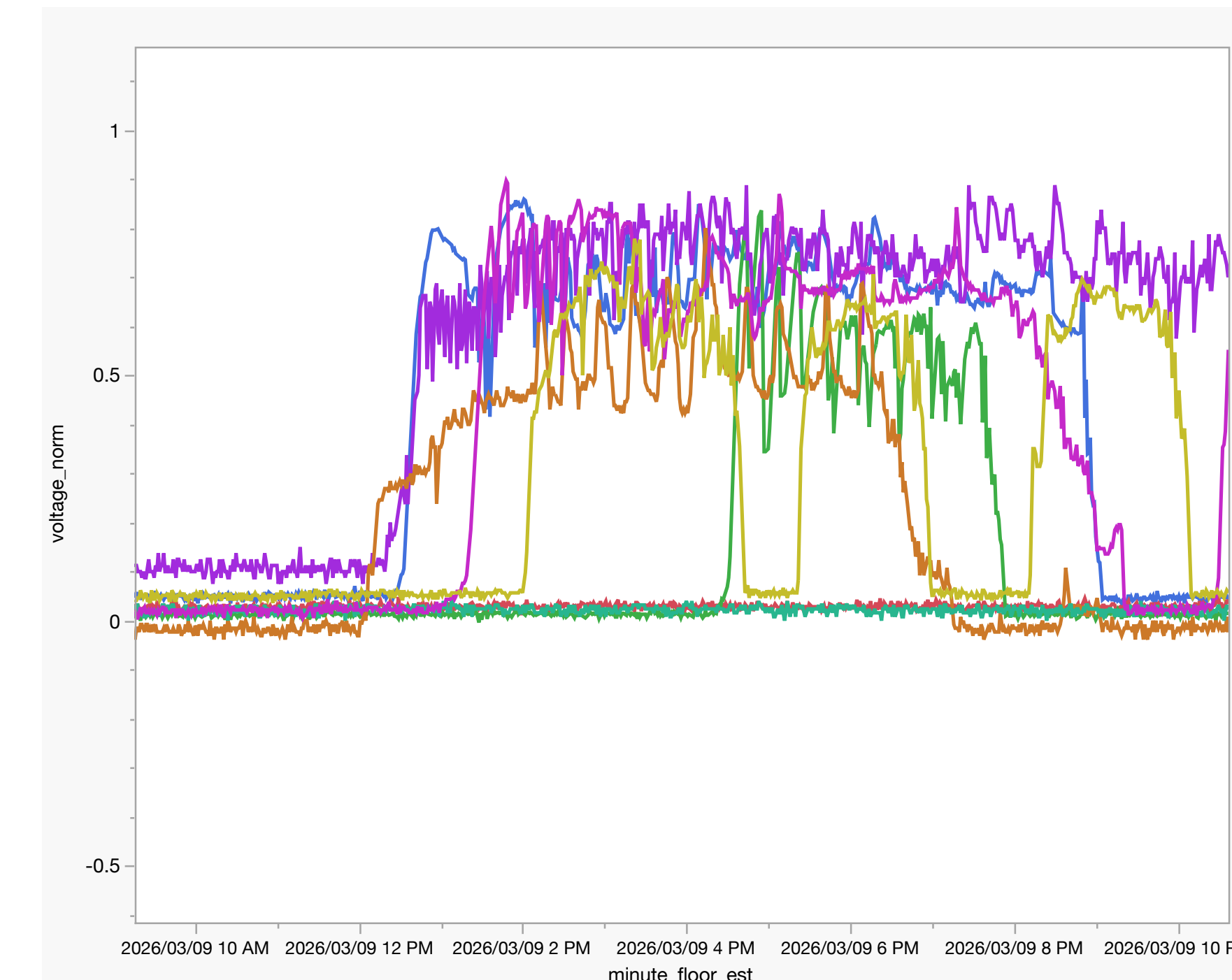


Figure 4: Oyster behavior in low salinity treatment after feeding at 12:15pm .

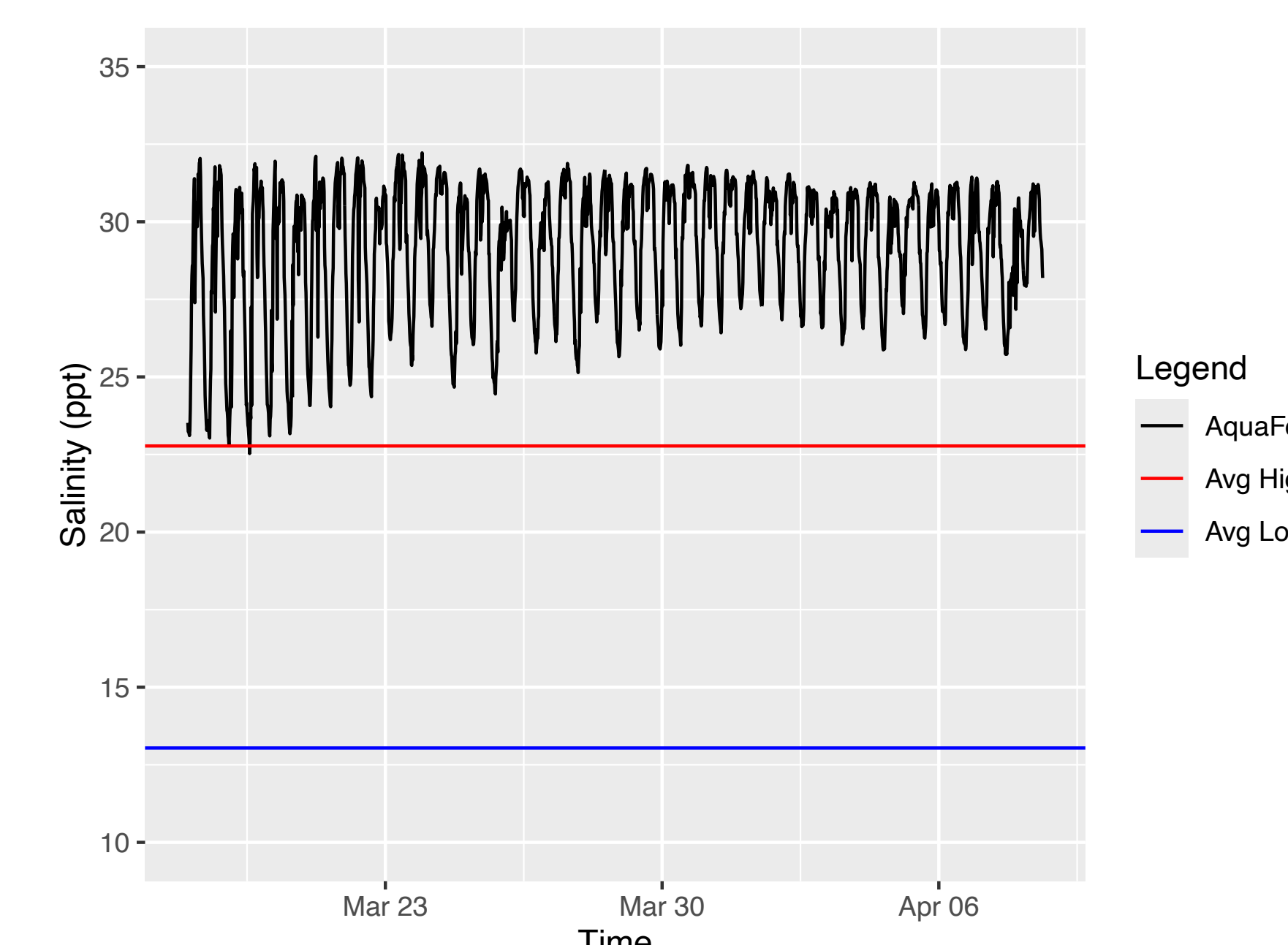


Figure 6: Salinity at Aquafort during biosensor deployment with average salinity values of high and low salinity lab treatments.

## Conclusion

- In the lab, under high salinity conditions, the oysters remained open much longer, indicating that low salinity could inhibit their physiological functions associated with gaping.
- Although the Aquafort had a similar salinity level compared to the high salinity lab treatment (~25ppt), the oysters did not open as frequently. This suggests that oysters are remaining closed due to a variable other than salinity.
- The biosensor unit could be utilized for future research into the gaping behaviors of oysters under various conditions.<sup>6</sup>
  - Temperature, pH, dissolved oxygen, salinity
  - Should include larger sample sizes.
- Deployments are now possible on many natural reefs, allowing for continuous site observation.
- Inexpensive biosensor systems can be used to monitor oyster success instead of expensive systems that monitor potential success metrics.

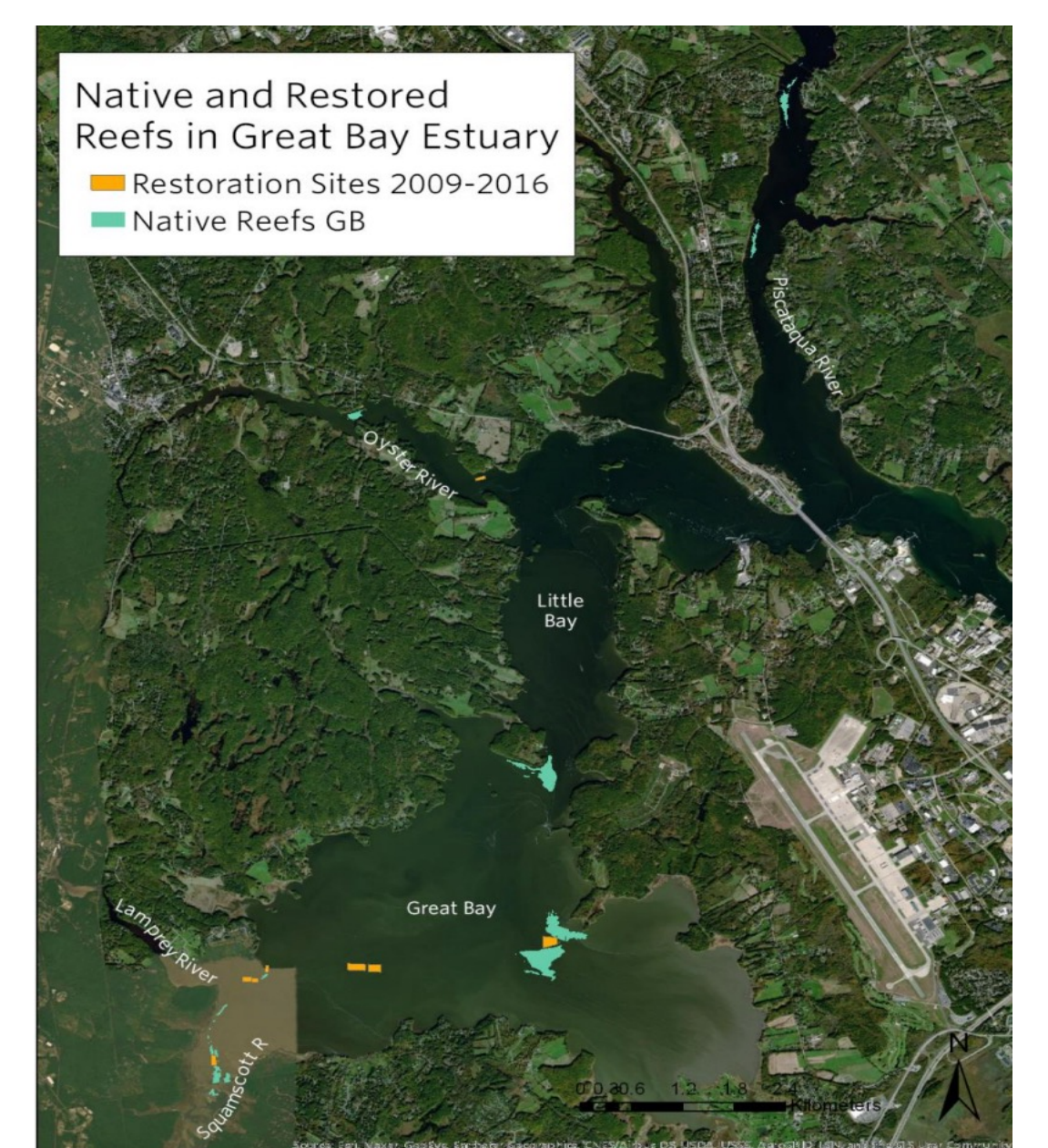


Figure 6: Map of native and restored oyster reefs in Great Bay<sup>1</sup>

## Acknowledgements

This work was funded in part by New Hampshire Sea Grant's Workforce Development Project E/WFD-3, pursuant to National Oceanic and Atmospheric Administration Award No. NA24OARX417C0037.

## References

1. Barr, J. M., Munroe, D., Rose, J. M., Calvo, L., Cheng, K. M., Bayer, S., & Kreeger, D. (2024). Seasonal feeding behavior of aquaculture eastern oysters (*Crassostrea virginica*) in the Mid-Atlantic. *Estuaries and Coasts*, 47(3), 789-804.
2. Casas, S. M., Filgueira, R., Lavaud, R., Comeau, L. A., La Peyre, M. K., & La Peyre, J. F. (2018). Combined effects of temperature and salinity on the physiology of two geographically-distant eastern oyster populations. *Journal of Experimental Marine Biology and Ecology*, 506, 82-90.
3. Casas, S. M., Comba, D., La Peyre, M. K., Rikard, S., & La Peyre, J. F. (2024). Rates of osmoconformation in triploid eastern oysters, and comparison to their diploid half-siblings. *Aquaculture*, 580, 740326.
4. Hoellein, T. J., Zarnoch, C. B., & Grizzle, R. E. (2015). Eastern oyster (*Crassostrea virginica*) filtration, biodeposition, and sediment nitrogen cycling at two oyster reefs with contrasting water quality in Great Bay Estuary (New Hampshire, USA). *Biogeochemistry*, 122(1), 113-129.
5. Laferriere, A., & group, B. (2021). *Restoration by design: Great Bay Estuary, New Hampshire*. (Report prepared for the U.S. Department of Agriculture Natural Resources Conservation Services). The Nature Conservancy. <https://www.nature.org/content/dam/tnc/nature/en/documents/RestorationByDesignNHFinalReport.pdf>
6. Yongue, S. M. (2025). *The Eastern Oyster Behavioral Response to Low Salinity and Hypoxia in Controlled and Natural Settings* (Master's thesis, Louisiana State University and Agricultural & Mechanical College).