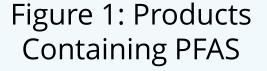


# Introduction

PFAS = Per- and Polyfluoroalkyl Substances

- Don't fully break down in the environment due to strength of carbon-fluorine bond
- Associated with negative health impacts including thyroid disease, cancer, and liver disease (CDC, 2024)





• PFAS are frequently detected in municipal wastewater, but little is known about the presence, concentrations, and breakdown mechanics of PFAS in septic systems

### Motivation

- Because PFAS are used in many commercial products like detergents and other cleaning agents, these compounds are being detected in household septic systems
- Studies have shown that septic systems are a non-point source of PFAS and other organic wastewater contaminants (OWCs) to groundwater (Schaider et al., 2016)
- Households that rely on septic systems for the treatment of wastewater as well as groundwater as a drinking water source are common in NH and especially vulnerable, as PFAS-containing septic discharge can migrate towards drinking water wells even with lengthy setback requirements (See Figure 2)

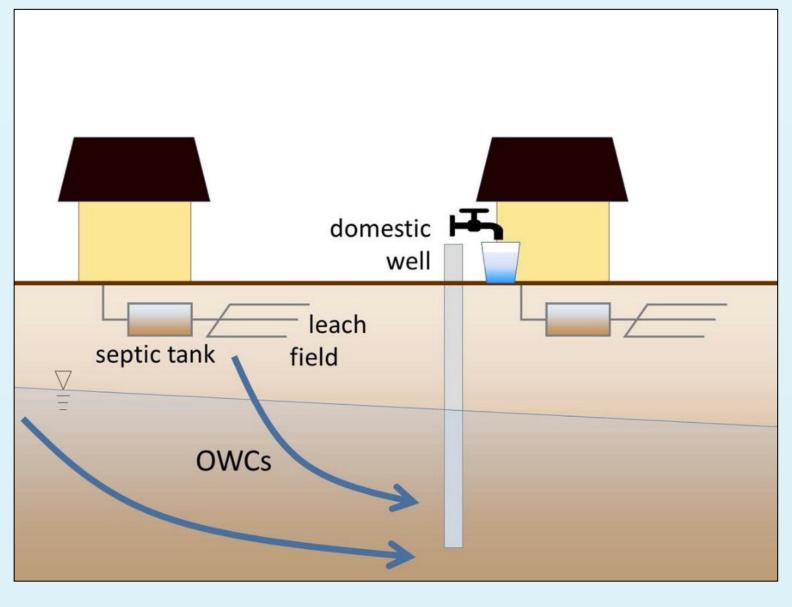
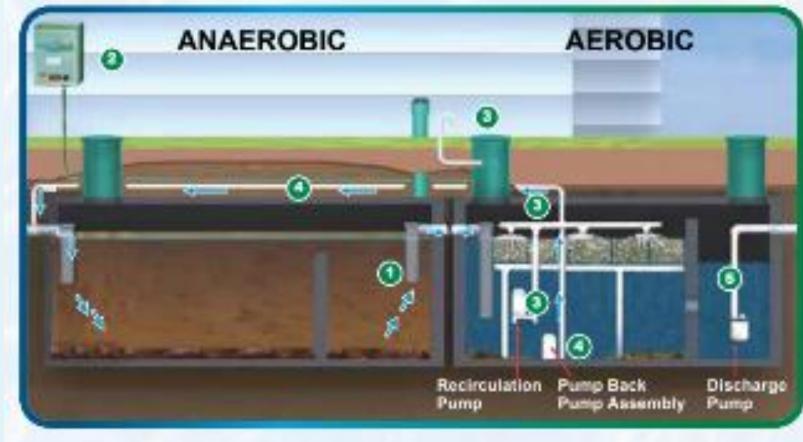
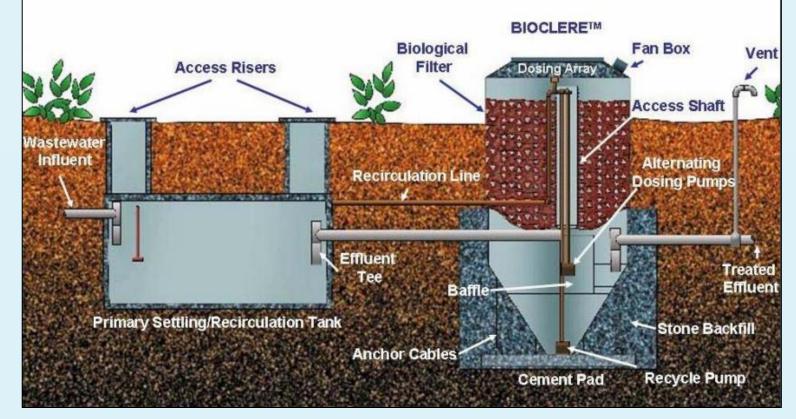


Figure 2: Septic Systems as a Non-point Source of PFAS to Drinking Water (Schaider et al., 2016)

- 1) Determine the types and typical concentrations of PFAS commonly found in septic systems in comparison to municipal wastewater treatment systems
- 2) Understand how septic systems designed for nitrification and denitrification processes influence PFAS biochemical transformations
- 3) Examine how PFAS partition between solids and liquids within septic systems
- 4) Identify the types of microbial communities present in septic systems associated with PFAS biotransformation
- 5) Determine the influence that environmental conditions such as temperature, dissolved oxygen concentration, conductivity, and pH have on PFAS biotransformation

denitrification systems





# Characterization of PFAS Occurrence, Transformation, and Partitioning in New Hampshire Community Septic Systems Alexis Eaton<sup>1</sup>, Jennifer Harfmann<sup>2</sup>, Stephen Roy<sup>2</sup>, James P. Malley<sup>1</sup>, and Paula J. Mouser<sup>1</sup>

<sup>1</sup>Civil and Environmental Engineering, University of New Hampshire, Durham <sup>2</sup>New Hampshire Department of Environmental Services

### **Project Goals**

The goal of this project is to identify gaps in current knowledge about PFAS in septic systems:

# Location of Research

- Sites to be sampled include two community septic systems in coastal New Hampshire that have nitrification/
- First site utilizes a SeptiTech STAAR system (Figure 3)

Figure 3: SeptiTech STAAR (Smart Trickling Aerobic/Anaerobic Recirculation) System

• Other site uses a Bioclere trickling filter over a clarifier (Figure 4)

Figure 4: Bioclere Trickling Filter

- collected semiannually from both sites
- Influent and effluent (liquid) samples collected monthly • Post-nitrification and post-denitrification samples
- collected in summer of 2024 during routine pump out will be collected from both sites in January 2025 and June 2025 and analyzed using EPA Method 1633 • DNA extraction will also be performed to identify the microbial communities associated with biotransformations occurring in each septic system
- Solids samples from Site 1 settling tank were already • Biofilm present on the trickling filter media (Figure 5)
- Biofilm removed via sonication and vortexing



- Figure 5: Biofilm Present on the Trickling Filter Media at Site 2 and Sonication Process concentration, conductivity, and pH will be collected
- Measurements of temperature, dissolved oxygen with each sampling event

# **Preliminary Findings**

### Table 1: Detection Frequency of PFAS in WWTP and Septic System Influent

|  | Compound | WWTP Detection<br>Frequency | Septic System Detection<br>Frequency |
|--|----------|-----------------------------|--------------------------------------|
|  | PFBA     | 81%                         | 50%                                  |
|  | PFBS     | 69%                         | 83%                                  |
|  | PFPeA    | 86%                         | 83%                                  |
|  | PFHxA    | 98%                         | 67%                                  |
|  | PFHxS    | 62%                         | 50%                                  |
|  | PFHpA    | 69%                         | 83%                                  |
|  | PFOA     | 81%                         | 67%                                  |
|  | PFOS     | 50%                         | 50%                                  |

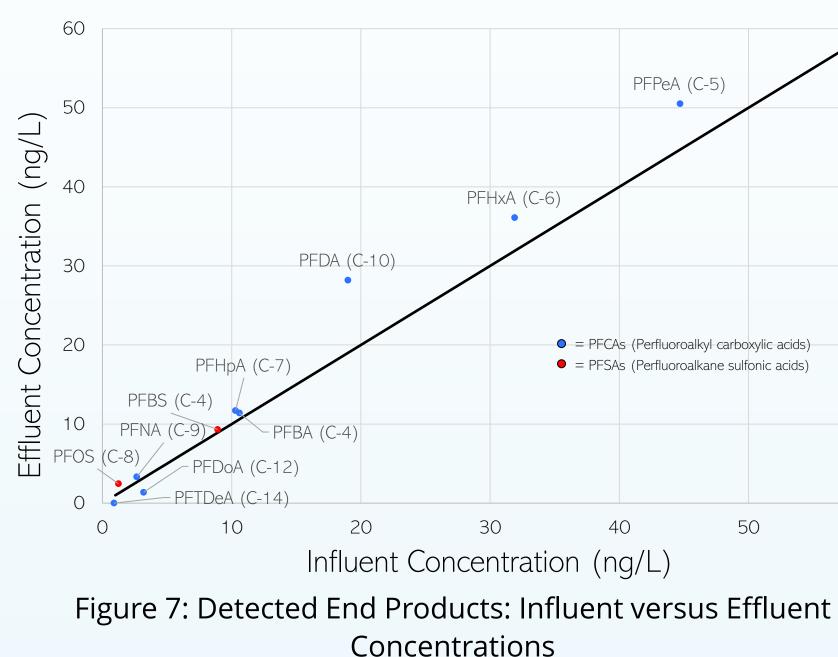
### Table 2: Detection Frequency of PFAS in WWTP and Septic System Effluent

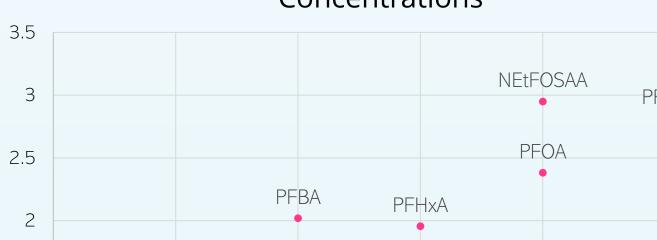
| Compound | WWTP Detection<br>Frequency | Septic System Detection<br>Frequency |
|----------|-----------------------------|--------------------------------------|
| PFBA     | 83%                         | 46%                                  |
| PFBS     | 86%                         | 67%                                  |
| PFPeA    | 100%                        | 83%                                  |
| PFHxA    | 100%                        | 100%                                 |
| PFHxS    | 60%                         | 25%                                  |
| PFHpA    | 88%                         | 75%                                  |
| PFOA     | 95%                         | 75%                                  |
| PFOS     | 85%                         | 58%                                  |



# Study Design

# **Preliminary Findings**





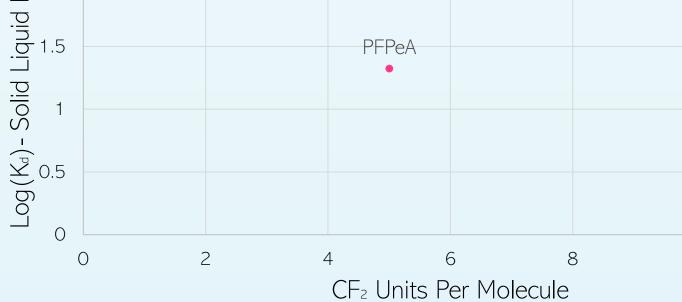


Figure 8: Solid-Liquid Partitioning in a Household Septic Tank Main Takeaways:

- Similar compounds present in municipal WWTFs are present in community septic systems
- Septic systems appear to be sources of short chain PFAS compounds and sinks for long chain compounds
- Higher concentrations of short chain PFAS in effluent than influent could be attributed to biotransformation
- It is likely that long chain PFAS are sorbing to the solids

### **Future Plans**

- Determine if preliminary findings and trends continue to be evident
- Perform DNA extraction of biofilm on media at both sites to characterize microbial communities associated with PFAS concentrations and biotransformation
- Examine how changes in environmental conditions over time influence PFAS concentrations (pH, temperature, etc.)
- Obtain post-nitrification and post-denitrification samples to understand how these processes influence PFAS transformation

